

An integrated approach for the effective
management of water pollution risks from
emerging contaminants



Perfluorinated compounds
HOlistic ENvironmental
Interinstitutional eXperience



Preventing. Ensuring. Promoting

LIFE PHOENIX Project

lifephoenix.eu

COORDINATOR



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AZIENDA
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UNIVERSITÀ
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DI PADOVA



WITH THE CONTRIBUTION OF THE LIFE FINANCIAL
INSTRUMENT OF THE EUROPEAN UNION
LIFE16ENV/IT/000488 - LIFE PHOENIX

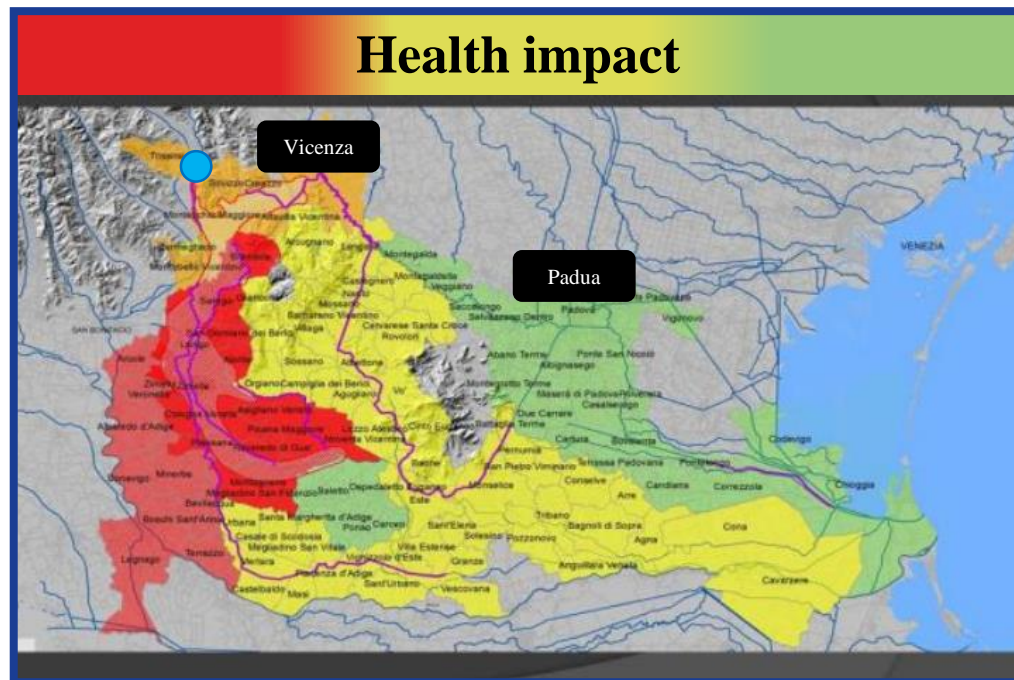
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Lesson learnt from three-years monitoring of irrigation waters, agricultural soil and plants

Claudia Ferrario
 IRSA-CNR Brugherio



Sampling area






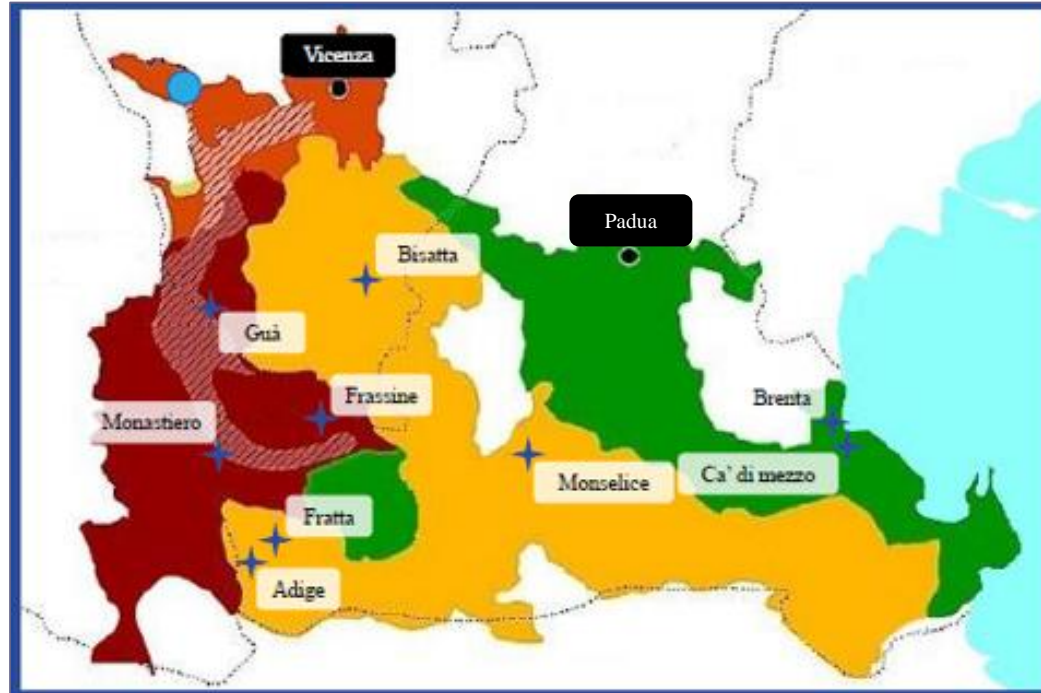
Matching the results of the biomonitoring study to those of drinking waters, Veneto local authority divided the contaminated area in three zones with different health impacts: zone of max exposure (red area), of precaution (yellow area) and not directly impacted (green area)

Veneto

Sampling sites



-  Contamination plume
-  Contamination source (fluorochemical industry)
-  Provincial borders



1 reference site
2 in green area
4 in yellow area
3 in red area

→ water body
used for irrigation →

agricultural soil
vegetal species
terrestrial invertebrates

Matrices

Abiotic



Biotic

Vegetables

- epigeal crop (lettuce)
- hypogeal crop (onion)
- extensive crop (maize)
- spontaneous plant (reed grass)



Animals



- terrestrial earthworms

Frequency of sampling

Abiotic



July
October



2018-2020

Biotic

Vegetables

- epigeal crop (lettuce)
- hypogeal crop (onion)
- extensive crop (maize)
- spontaneous plant (reed grass)



Animals



- terrestrial earthworms

Sampling size

Abiotic



2018-2020

Biotic

Vegetables

- epigeal crop (lettuce)
- hypogeal crop (onion)
- extensive crop (maize)
- spontaneous plant (reed grass)

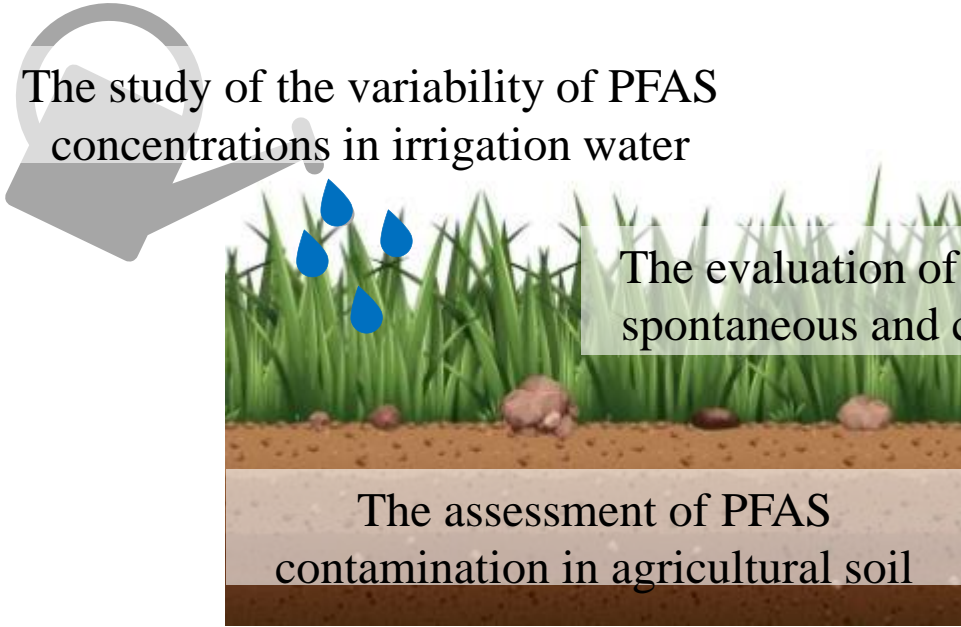


Animals



- terrestrial earthworms

Aims



The study of the variability of PFAS concentrations in irrigation water

The evaluation of PFAS uptake in spontaneous and cultivated plants

The assessment of PFAS contamination in agricultural soil

Analytical procedures



Standard Operating Procedures (SOPs) for the analytical determination and quantification of PFAS in environmental matrices

[Deliverable n.1 of the Action C.1]

The analytical procedures were developed to optimize the determination of short-chain PFAS (C<8) due to their higher potential to be translocated and bioaccumulated in plants than long-chain congeners.

Project reference: LIFE16 ENV/IT/000488
 Project acronym: LIFE PHOENIX
 Project full title: Perfluorinated compounds Holistic Environmental Interinstitutional eXperience
 Action C.1: Environmental Monitoring
 Due date of deliverable: 30/09/2018
 Lead beneficiaries: ARPAV, CNR-IRSA
 Authors: Roberto Lava, Caterina Cecchinato, Massimiliano Prenzato, Fabio Biancato, Francesca Zanon e Michele Gerotto (ARPAV)
 Claudia Ferrario, Sara Valsecchi e Stefano Polesello (CNR-IRSA)

Dissemination Level

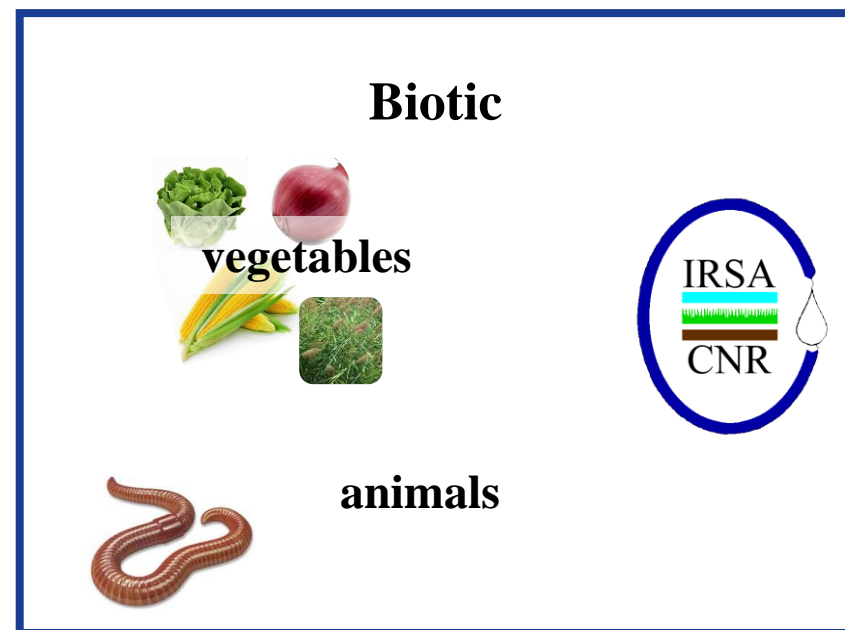
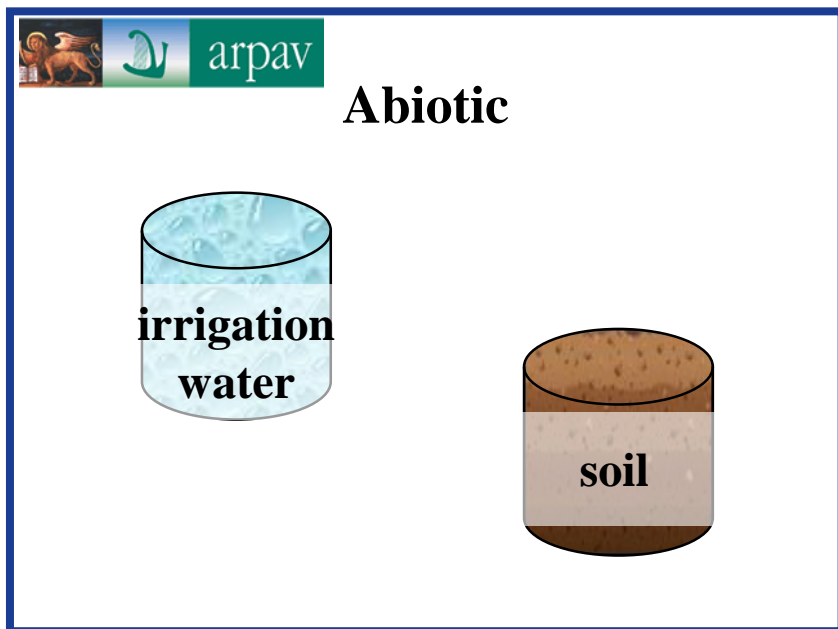
PU	Public	
CO	Confidential, only for members of the consortium (including the Commission Services)	X
CI	Classified, as referred to in Commission Decision 2001/844/EC	



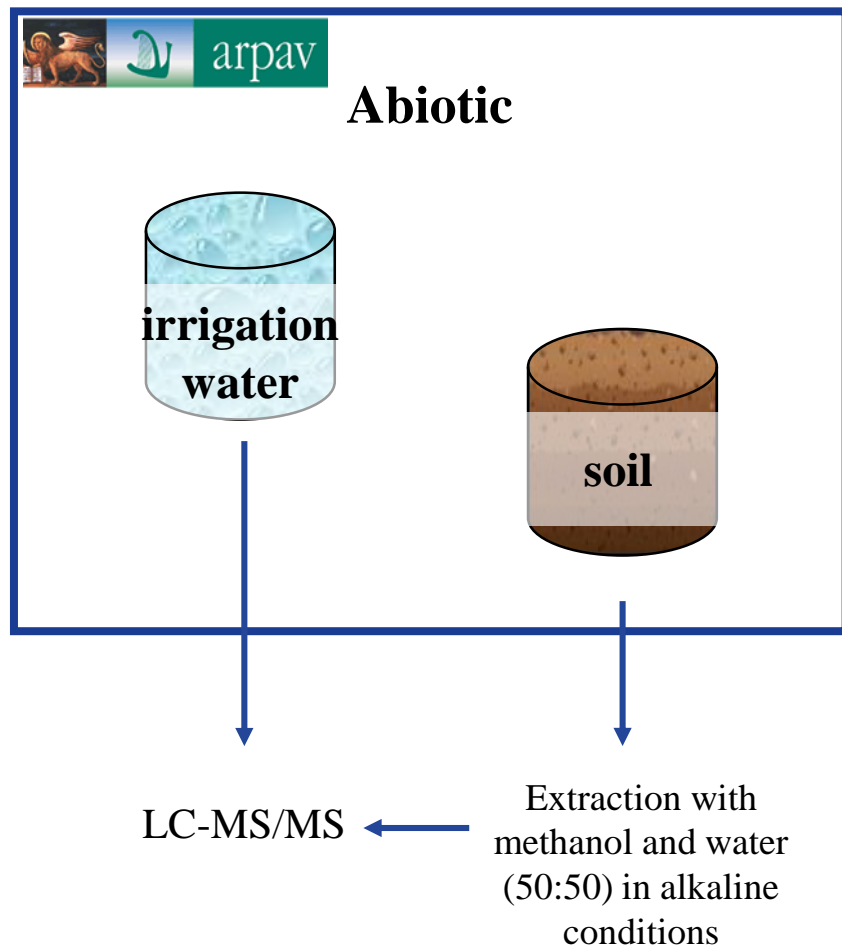
PFAS considered

Acido perfluorobutanoico	PFBA
Acido perfluoropentanoico	PFPeA
Acido perfluoroesanoico	PFHxA
Acido perfluoroeptanoico	PFHpA
Acido perfluorooottanoico	PFOA
Acido perfluorononanoico	PFNA
Acido perfluorodecanoico	PFDA
Acido perfluoroundecanoico	PFUnDA
Acido perfluorododecanoico	PFDoDA
Acido perfluorobutansolfonico	PFBS
Acido perfluoroesansolfonico	PFHxS
Acido perfluorooottansolfonico	PFOS

Analytical procedures



Analytical procedures

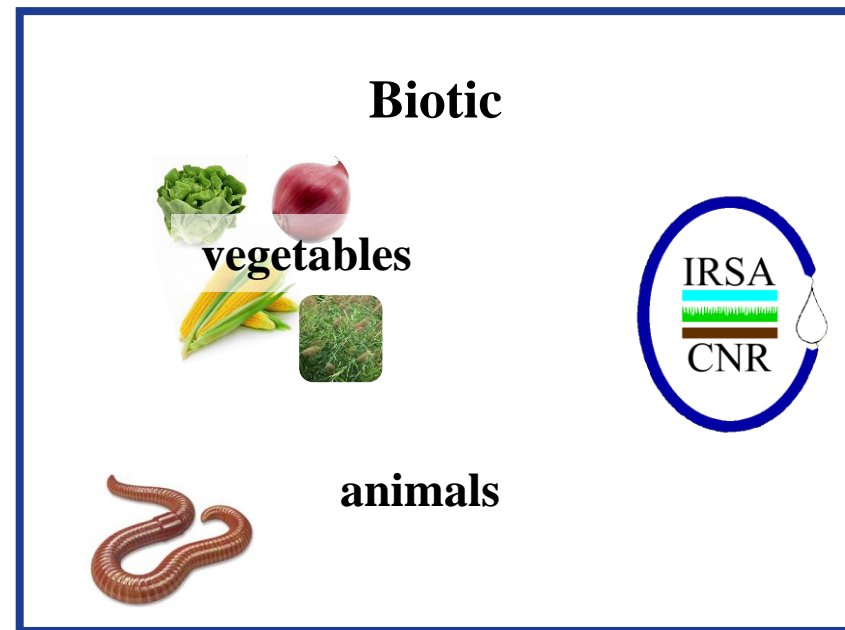
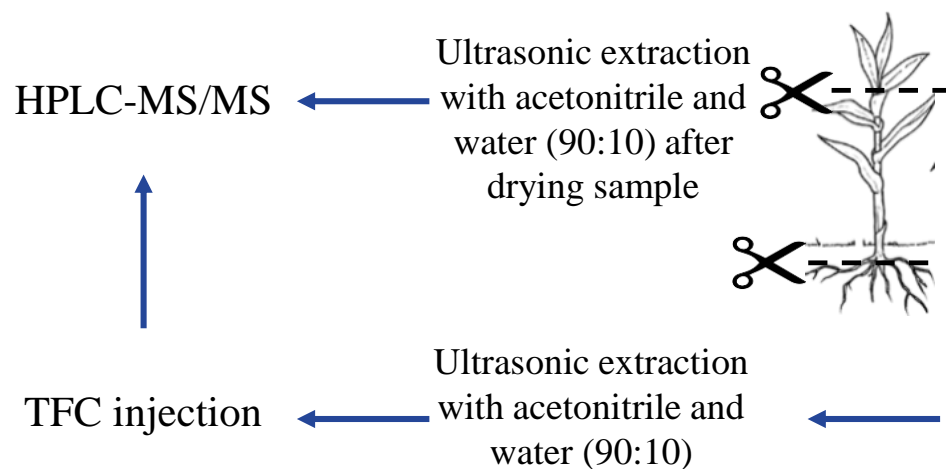


Analytical procedures

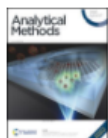
Webinar LIFE PHOENIX: metodiche per l'analisi di PFAS e contaminanti emergenti.

6 ottobre 2020

www.lifephoenix.eu/web/phoenix/-/ottimo-riscontro-per-il-webinar-pfas-e-contaminanti-emergenti



Analytical procedures



From the journal:
Analytical Methods

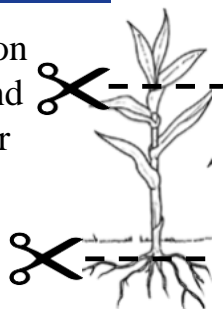
2021

Determination of perfluoroalkyl acids in different tissues of graminaceous plants

[CLAUDIA FERRARIO](#), [Sara Valsecchi](#), [Roberto Lava](#), [Marco Bonato](#) and [Stefano Polesello](#)

HPLC-MS/MS

← Ultrasonic extraction
with acetonitrile and
water (90:10) after
drying sample



Biotic

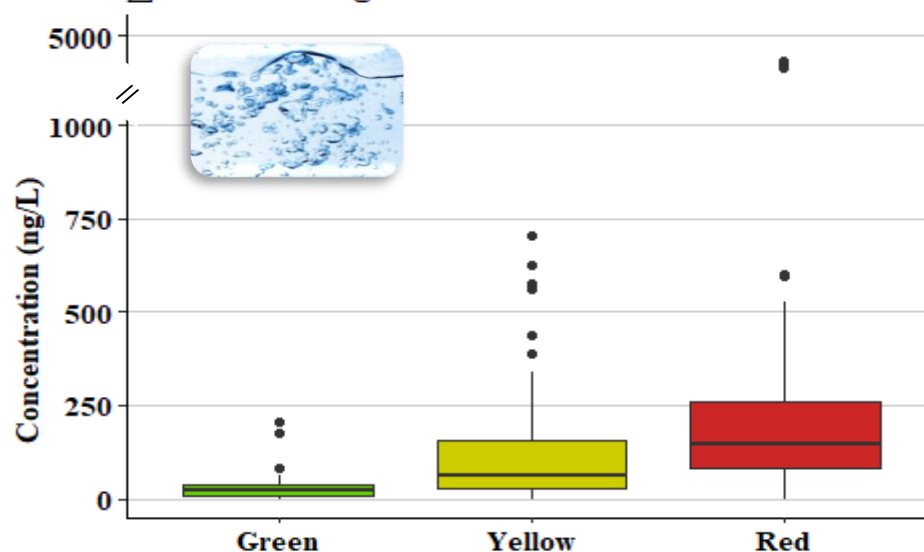


Results 2018-2020

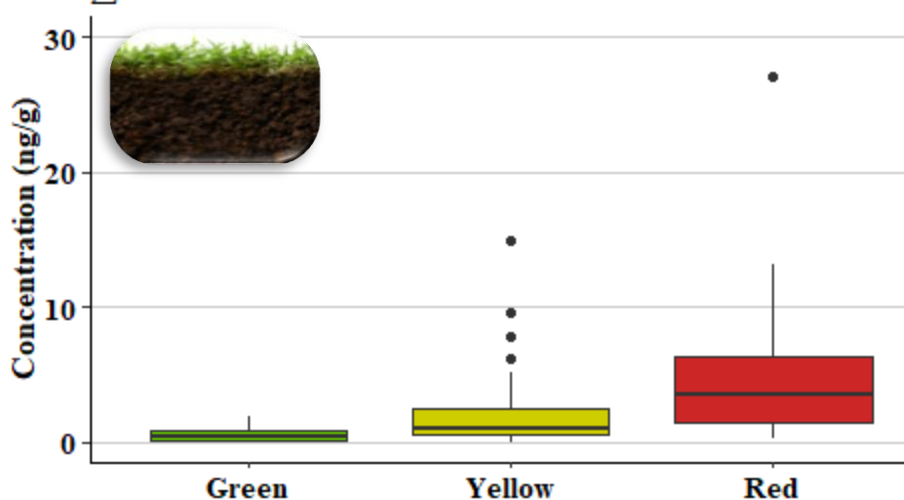
min-max
(average)

	Green	Yellow	Red
Water (ng/L)	<LOD – 217 (27)	<LOD – 764 (123)	<LOD – 4,694 (330)
Soil (ng/g)	<LOD – 1.9 (0.6)	<LOD – 14.9 (2.1)	<LOD – 27.1 (4.8)
Plant (ng/g ww)	<LOD – 4.8 (1.2)	<LOD – 9.8 (1.4)	<LOD – 10.4 (2)

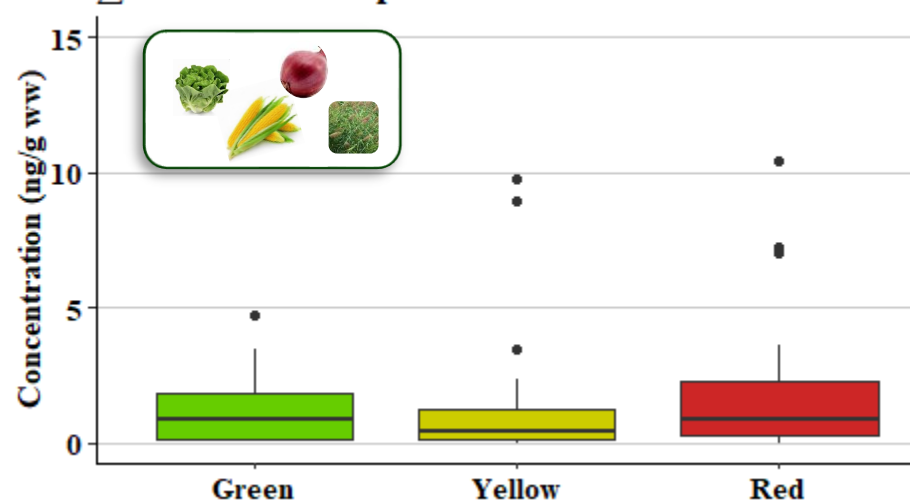
ΣPFAS in irrigation water



ΣPFAS in soil

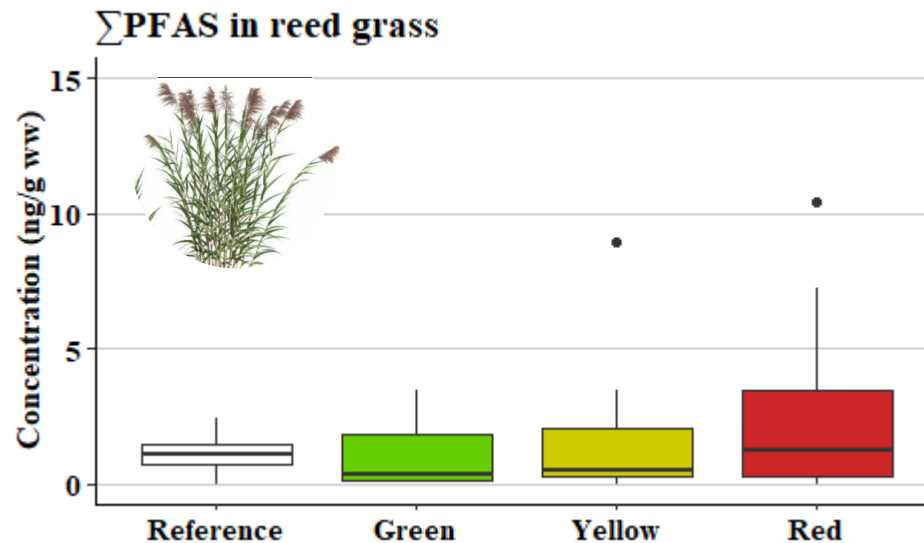
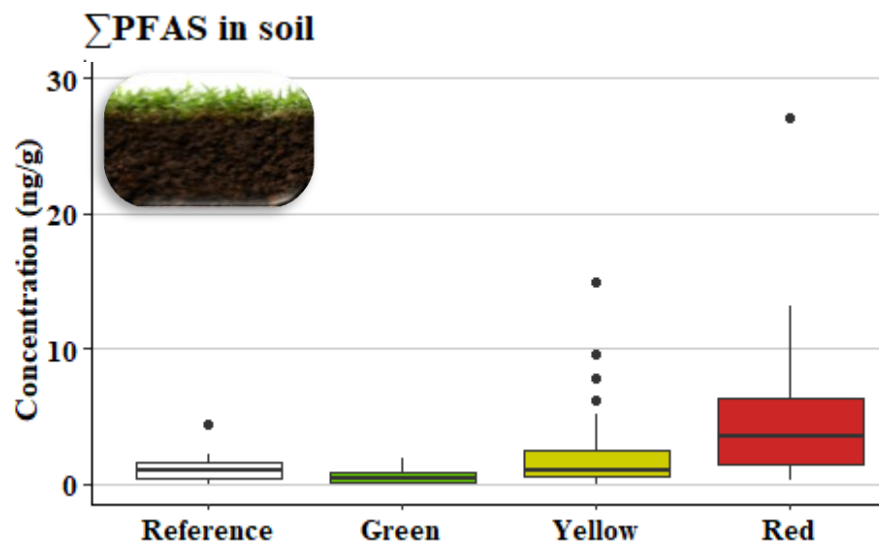
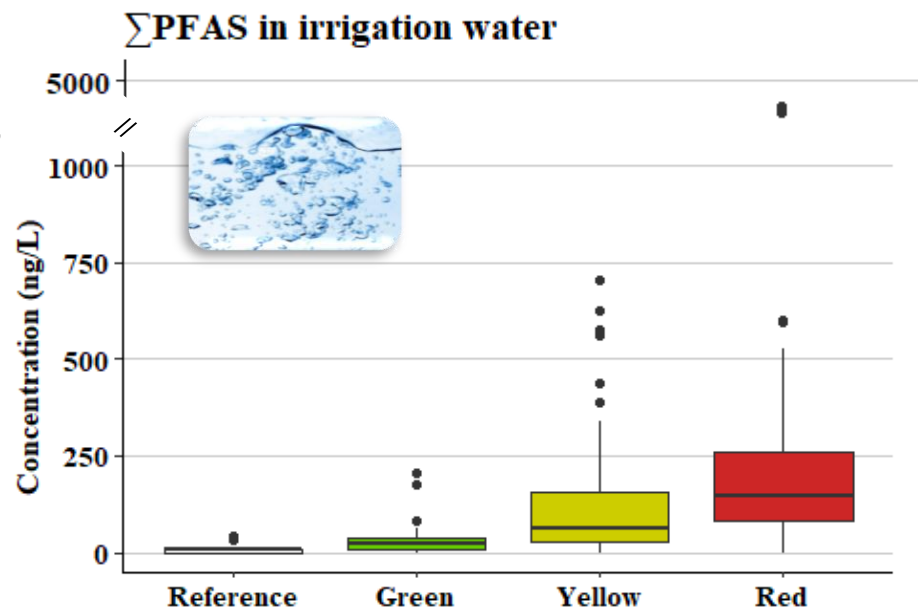


ΣPFAS in whole plant



Results 2018-2020

Comparison to “reference site”

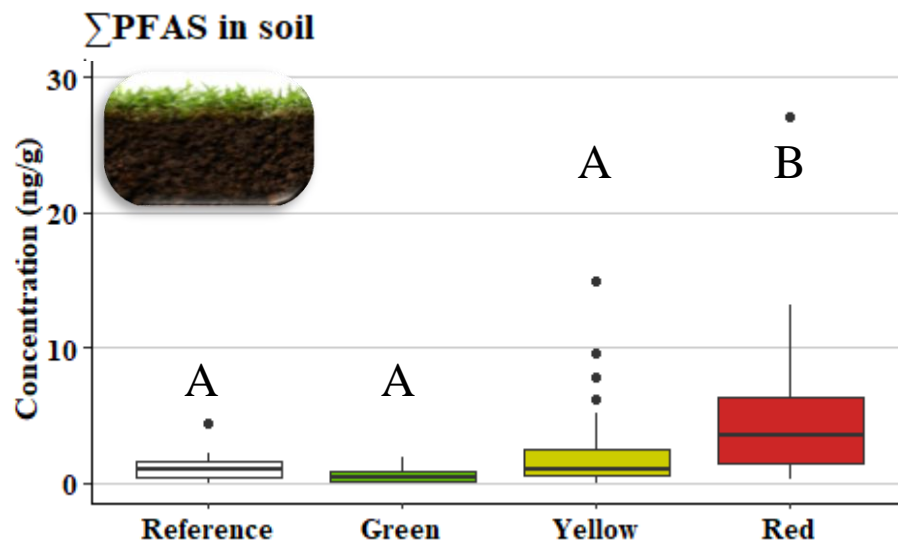
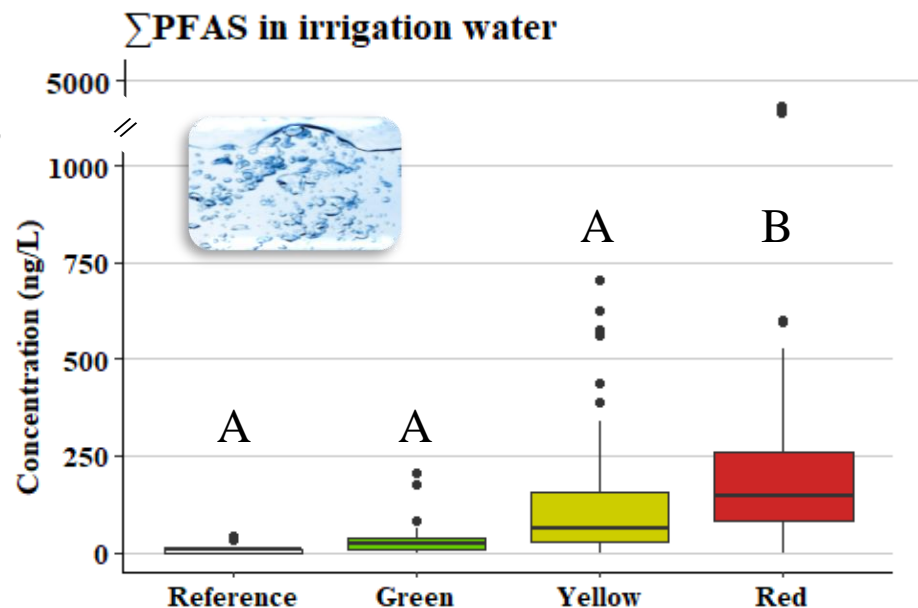


Results 2018-2020

Comparison to “reference site”

ANOVA test

PFAS concentrations in the red area were significantly higher than in other areas for waters and soil

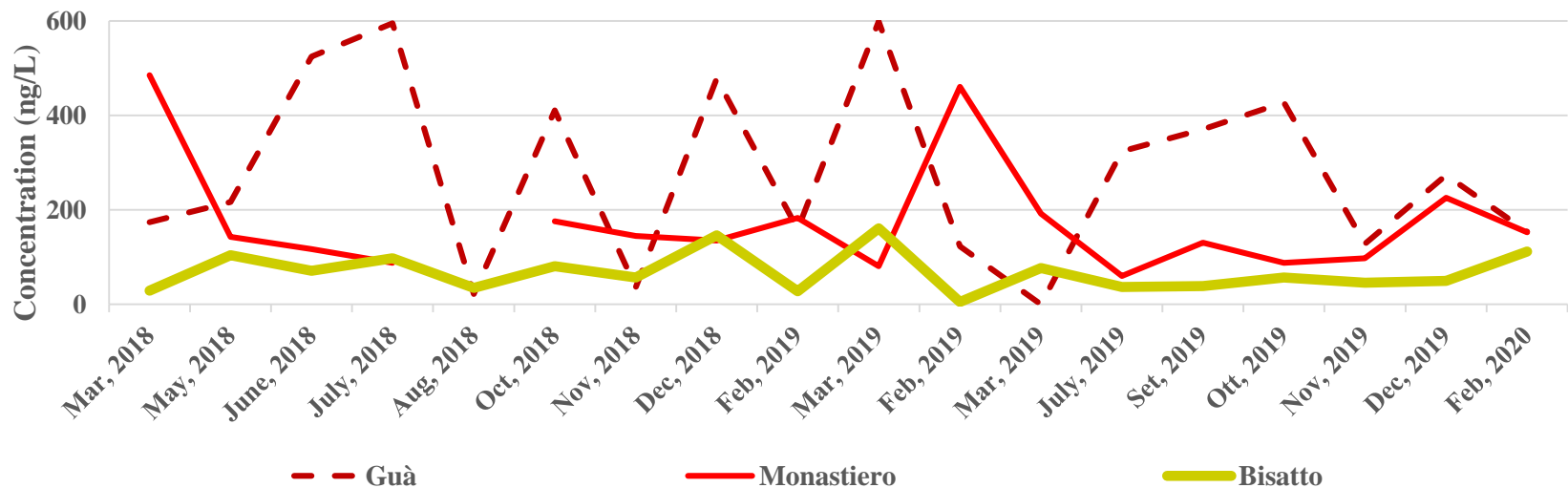
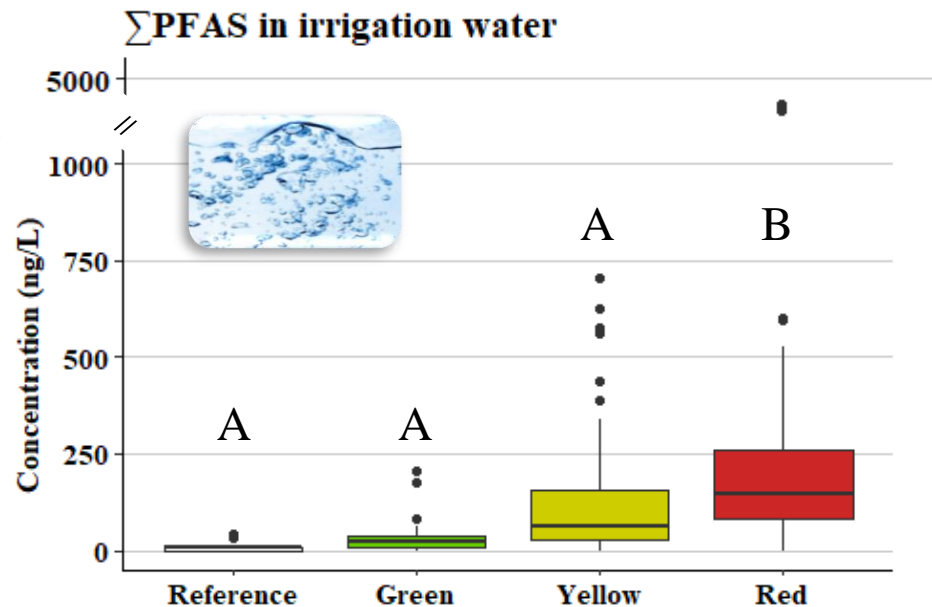


Green and yellow areas were no more contaminated by PFAS than the reference area

Results 2018-2020

Comparison to “reference site”

The levels of PFAS in irrigation waters showed high variability throughout the year



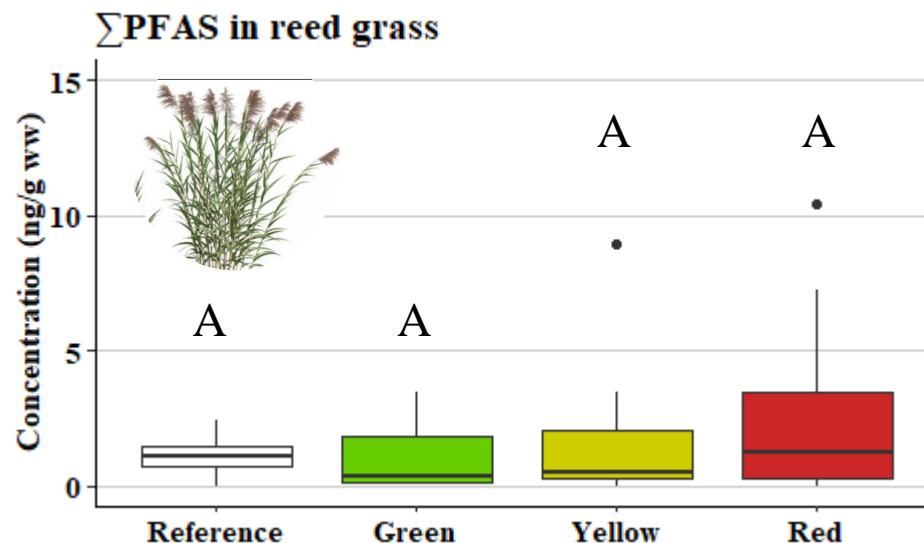
Results 2018-2020

Comparison to “reference site”



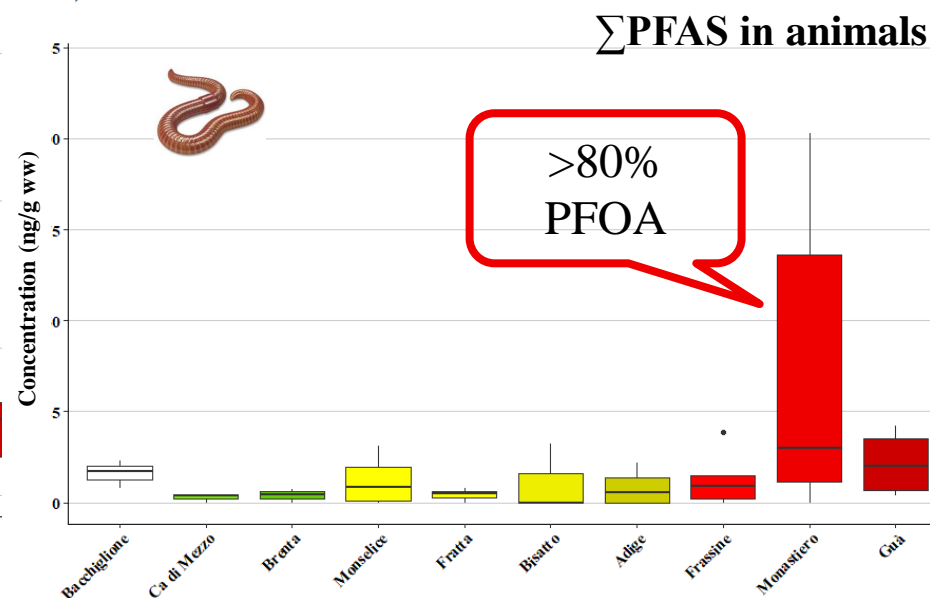
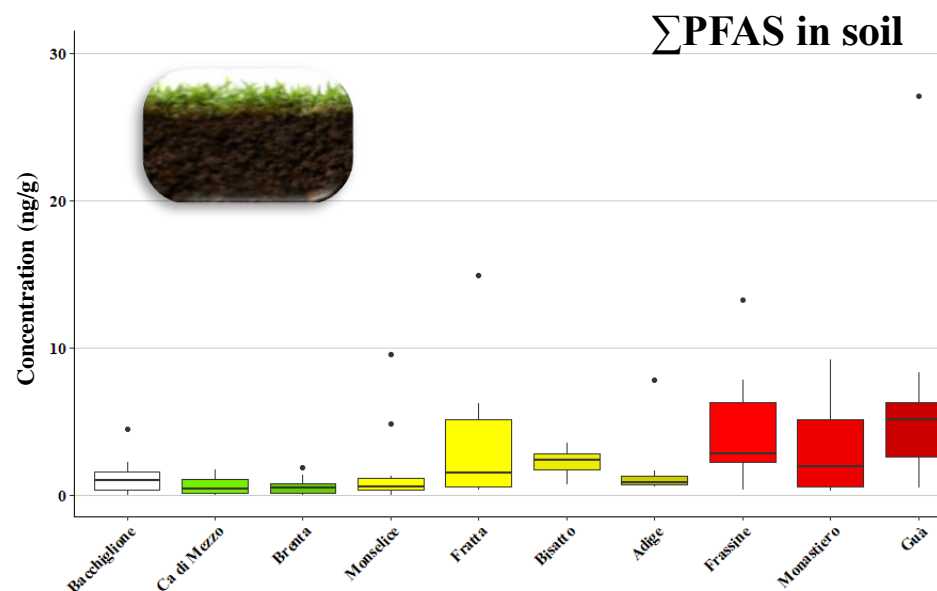
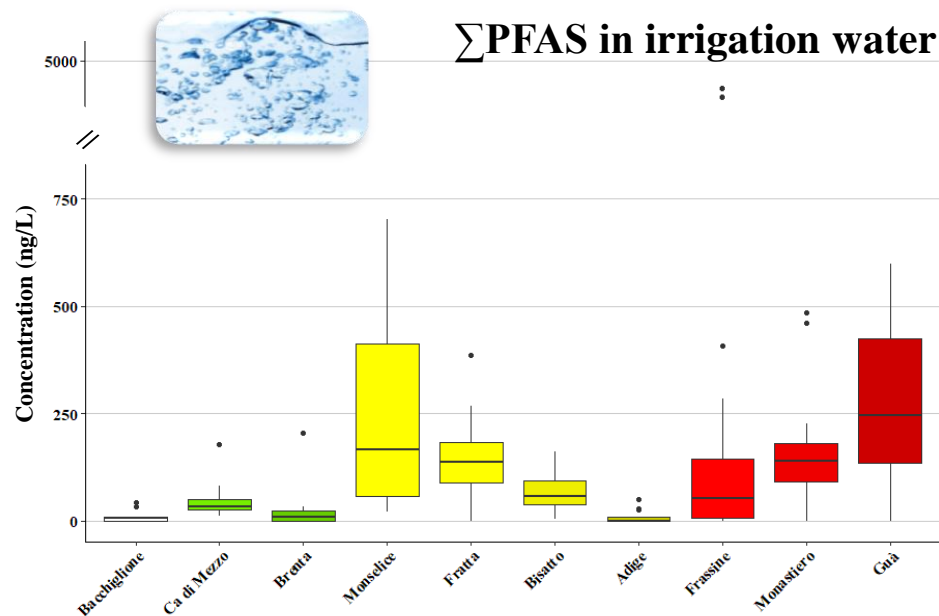
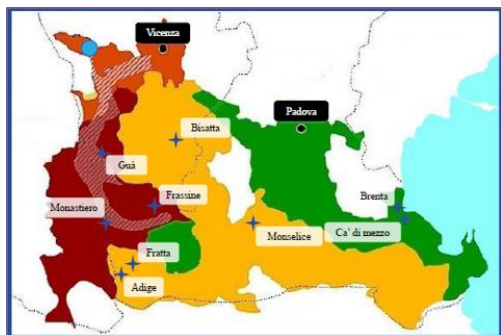
The three areas were no more contaminated by PFAS than the reference area

Reed grass is a ubiquitous aquatic vegetal species often used for phytodepuration in artificial wetlands



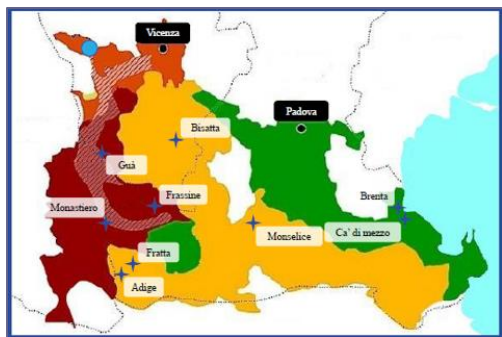
Results 2018-2020

Site comparison

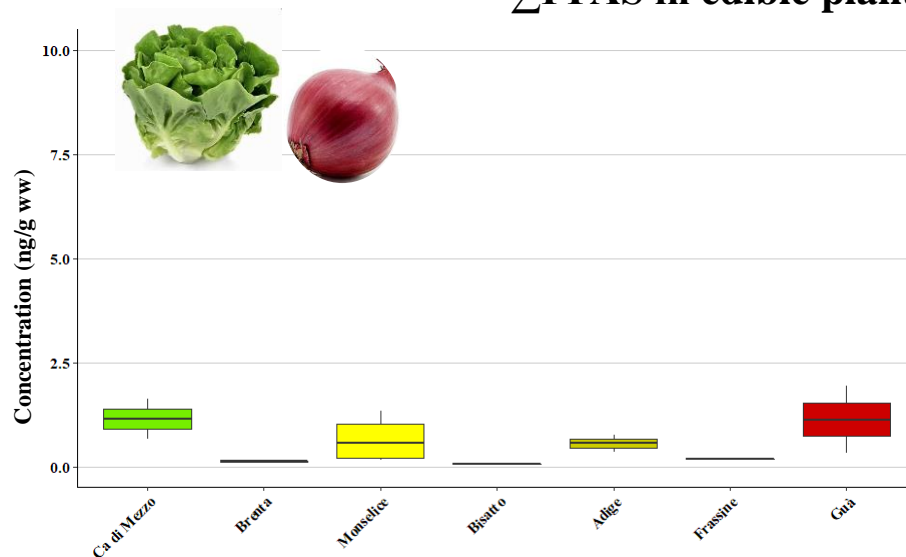


Results 2018-2020

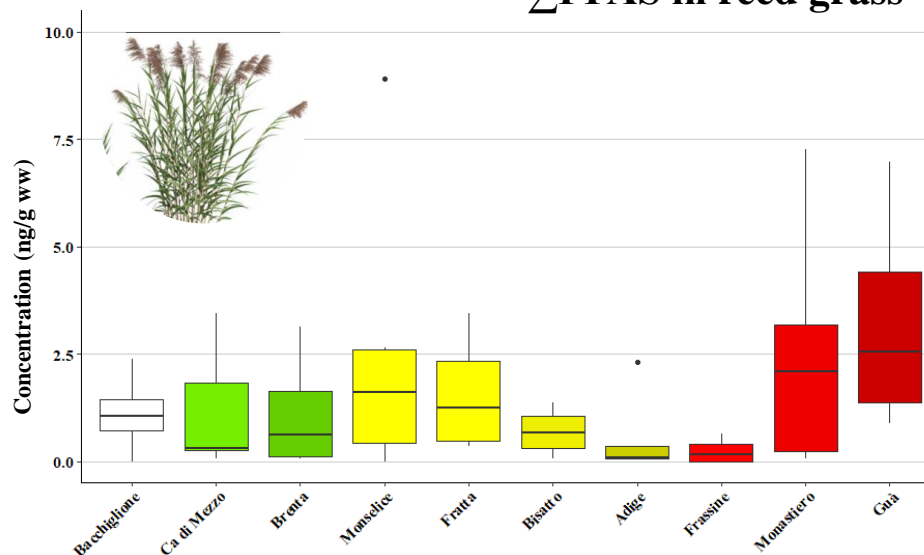
Site comparison



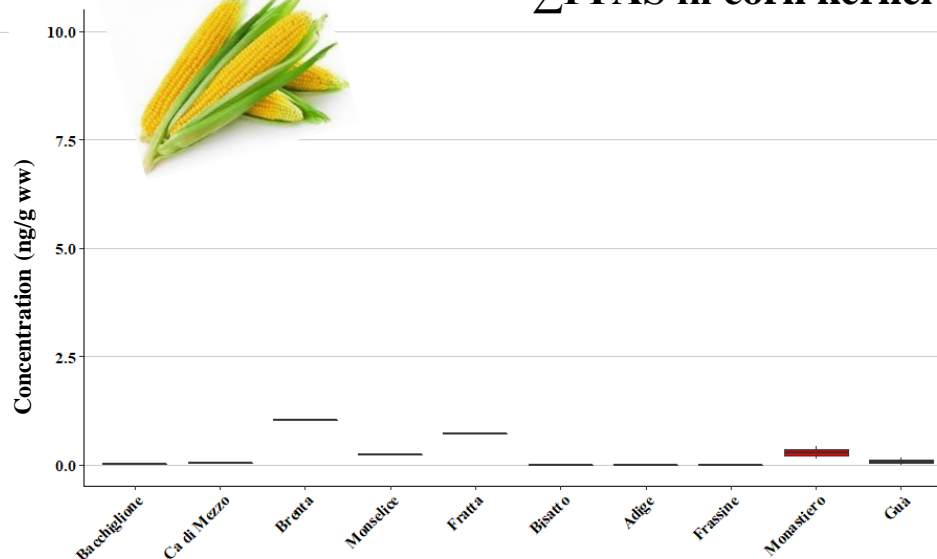
Σ PFAS in edible plants



Σ PFAS in reed grass



Σ PFAS in corn kernel



Results 2018-2020

PFAS composition

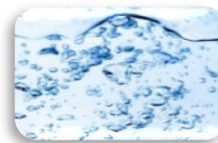
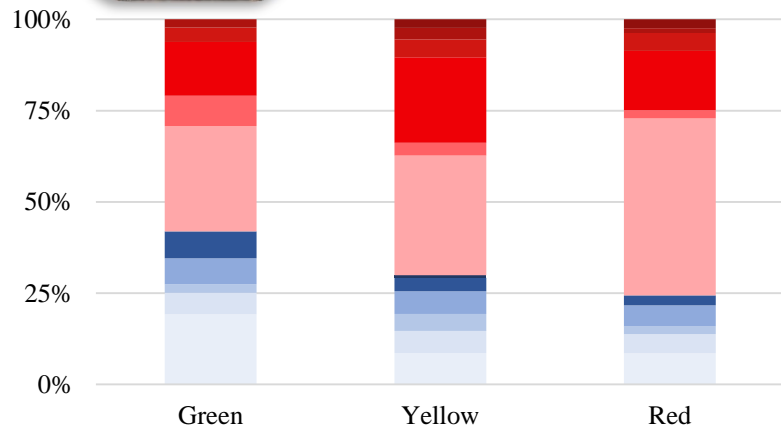
PFBA PFPeA PFBS PFHxA PFHpA PFHxS

■ C < 8 ■ C ≥ 8

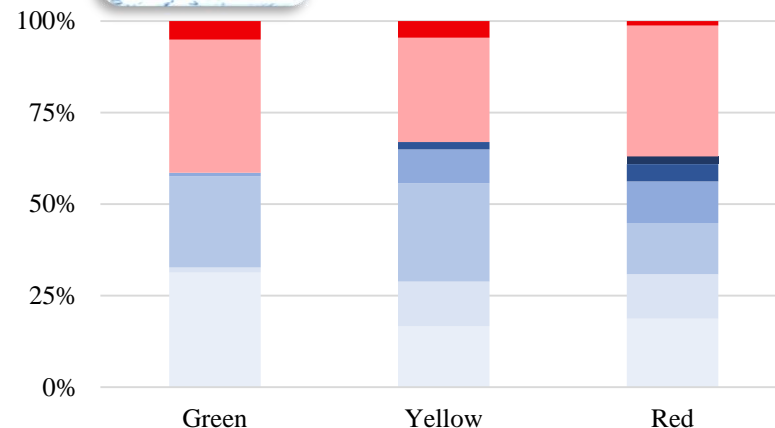
PFOA PFNA PFOS PFDA PFUnDA PFDODA



ΣPFAS in soil

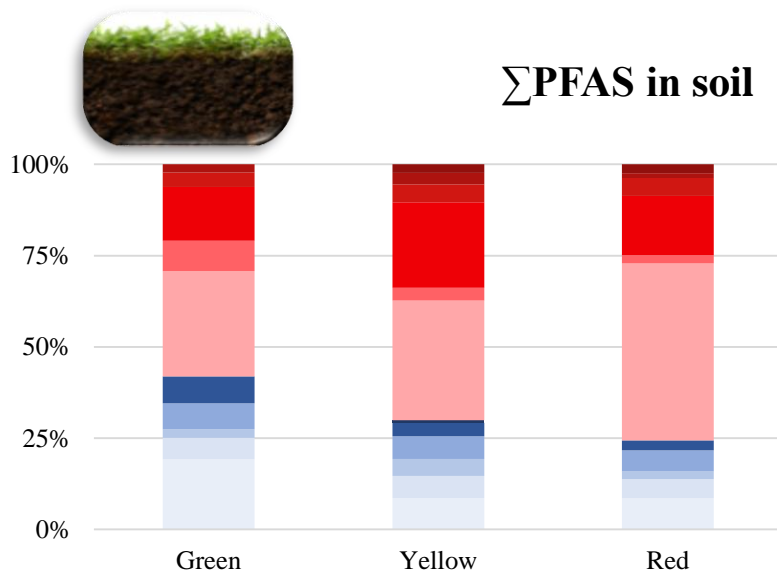
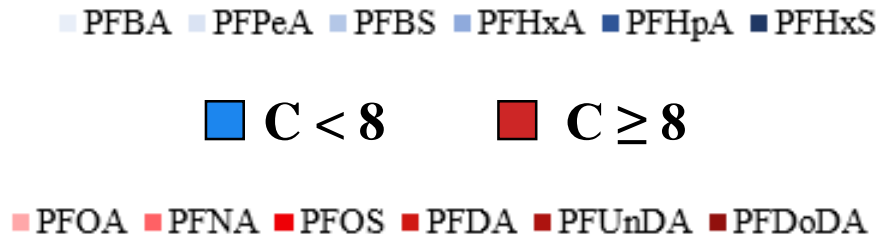


ΣPFAS in irrigation water

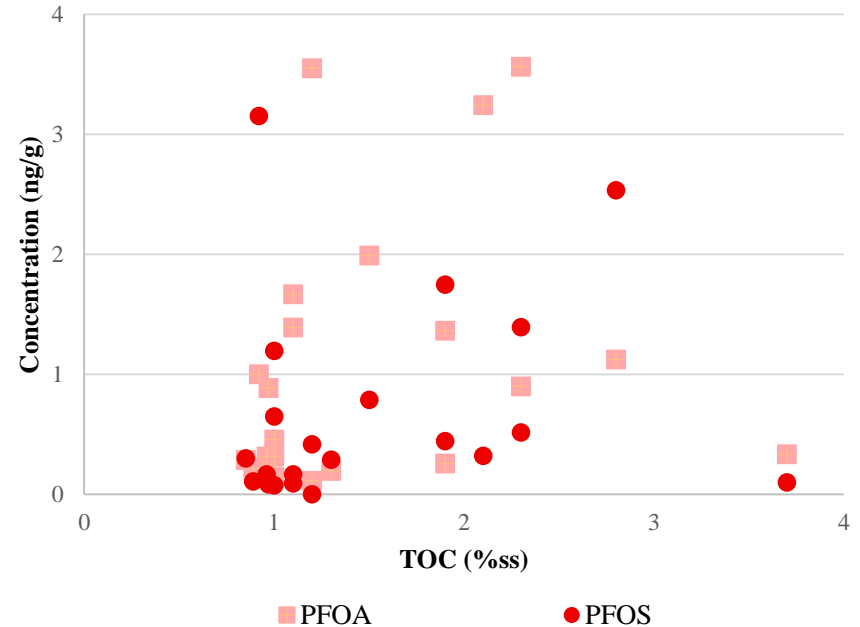


Results 2018-2020

Total organic carbon (TOC) in soil



TOC vs Σ PFAS in soil



No correlation between
TOC and Σ PFAS was
found

Results 2018-2020

PFAS composition

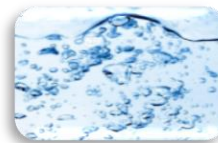
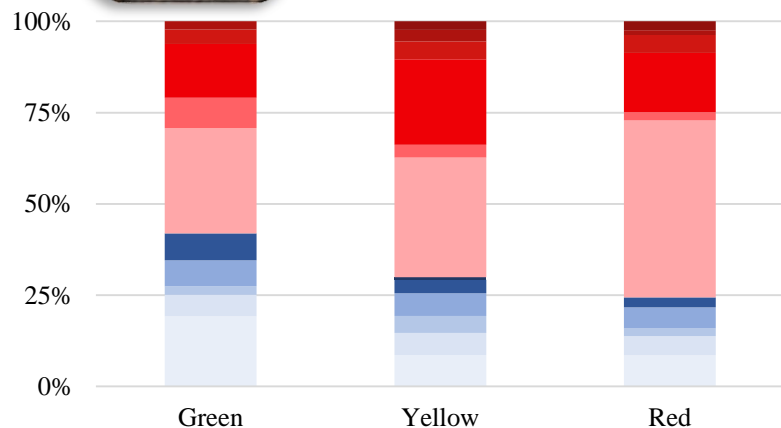
PFBA PFPeA PFBS PFHxA PFHpA PFHxS

$C < 8$ $C \geq 8$

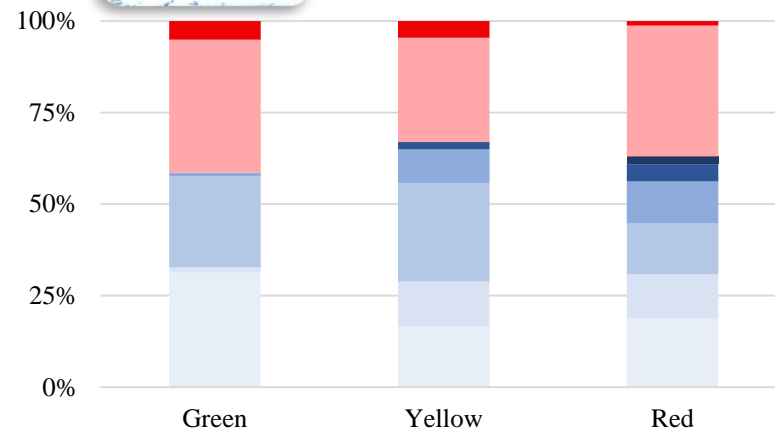
PFOA PFNA PFOS PFDA PFUnDA PFDODA



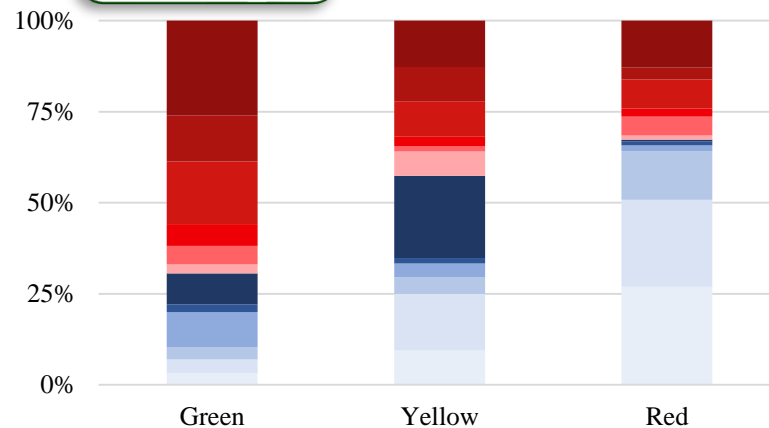
Σ PFAS in soil



Σ PFAS in irrigation water

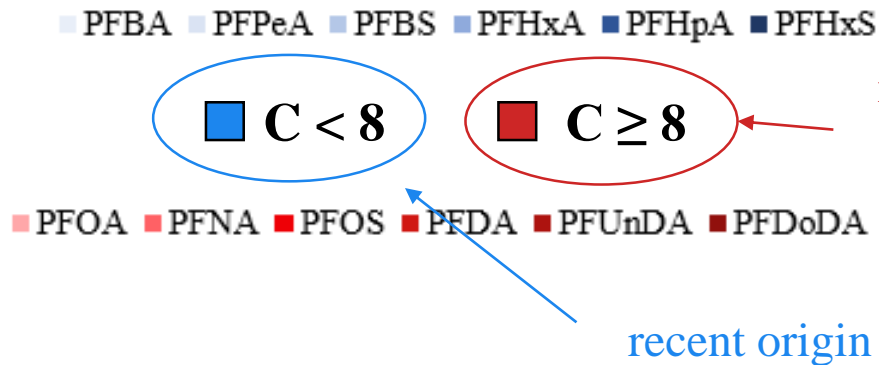


Σ PFAS in whole plant

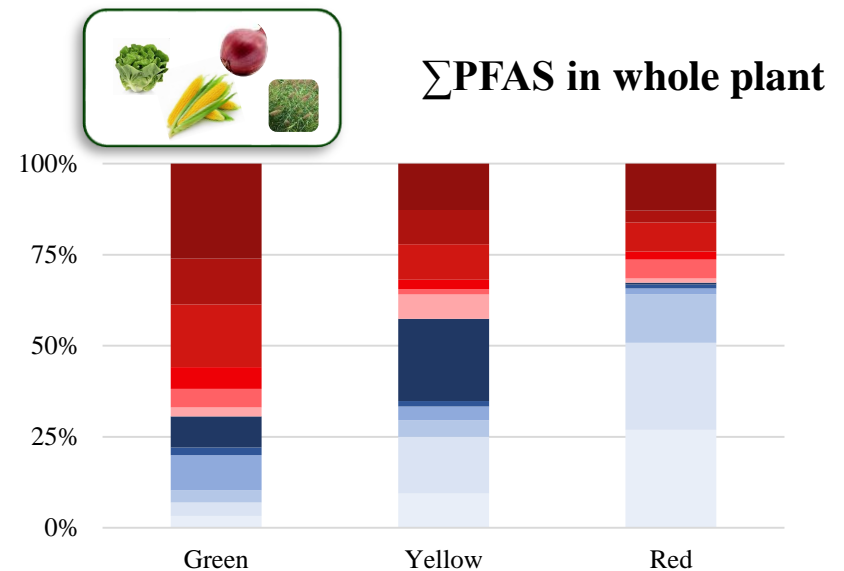


Results 2018-2020

PFAS composition in plants

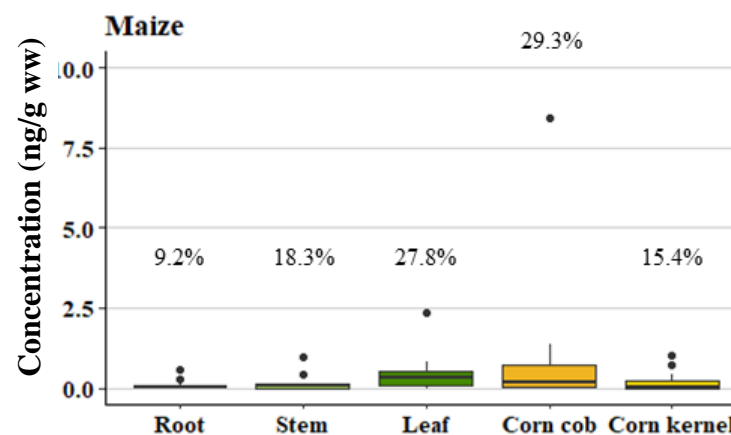
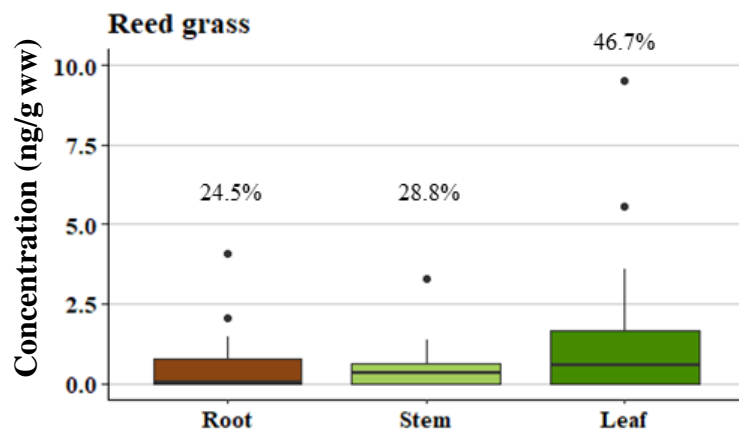
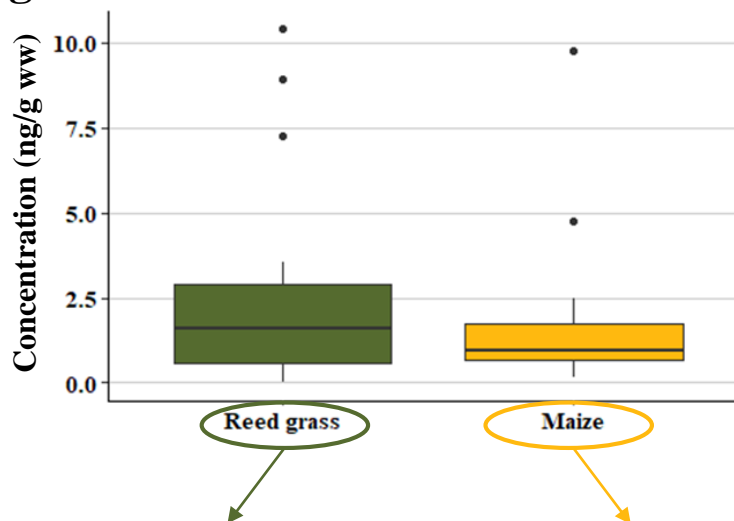


The composition of vegetal contamination was affected by the sampling site.



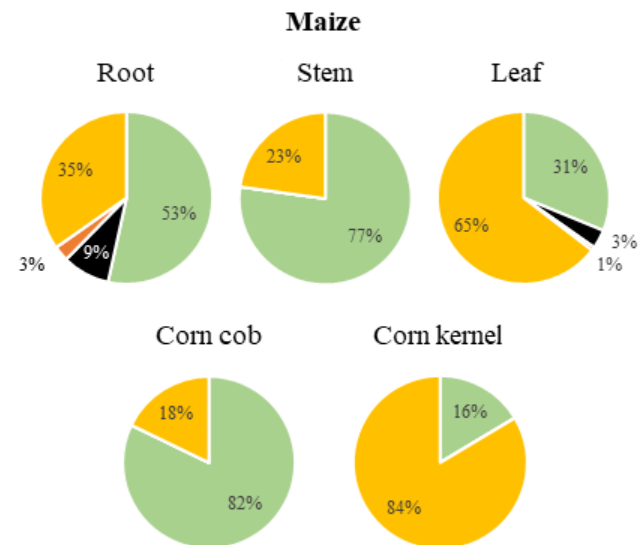
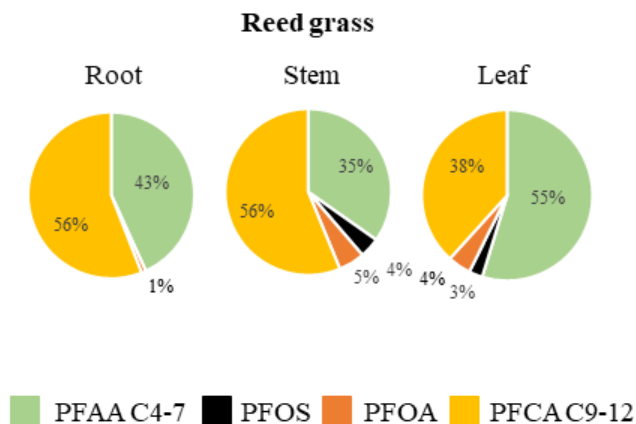
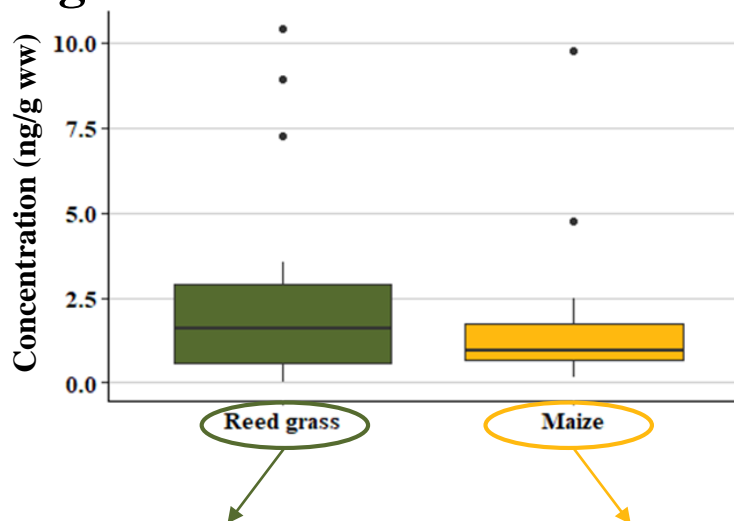
Results 2018-2020

PFAS partition in vegetal fractions

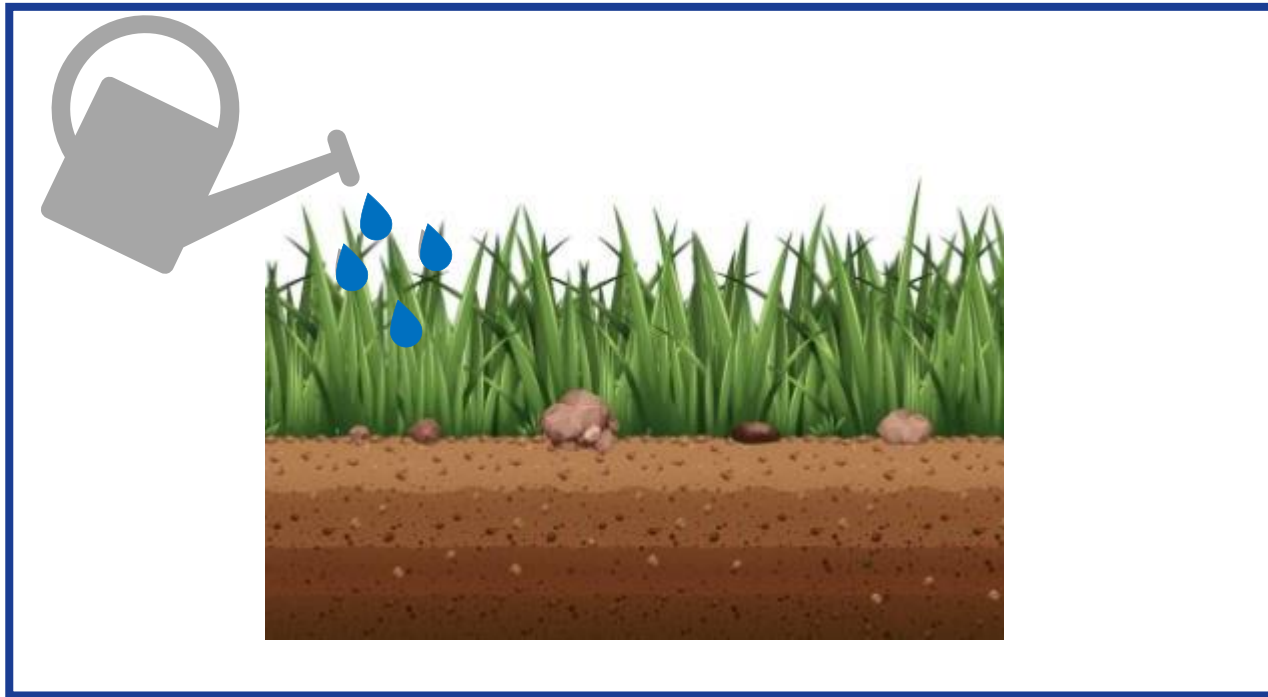


Results 2018-2020

PFAS composition in vegetal fractions



Conclusions



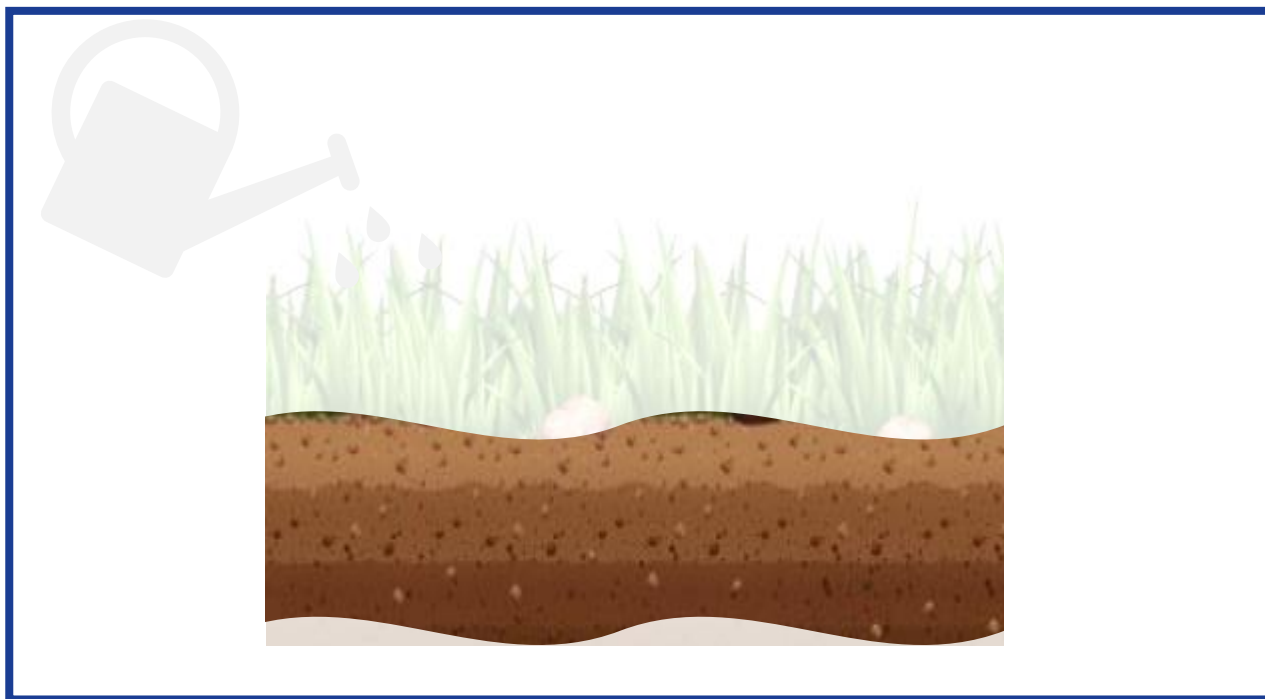
Conclusions

- The levels of PFAS in irrigation waters are characterized by a high variability due to the different water supply during the year.
- Events of higher contamination were occasionally recorded.



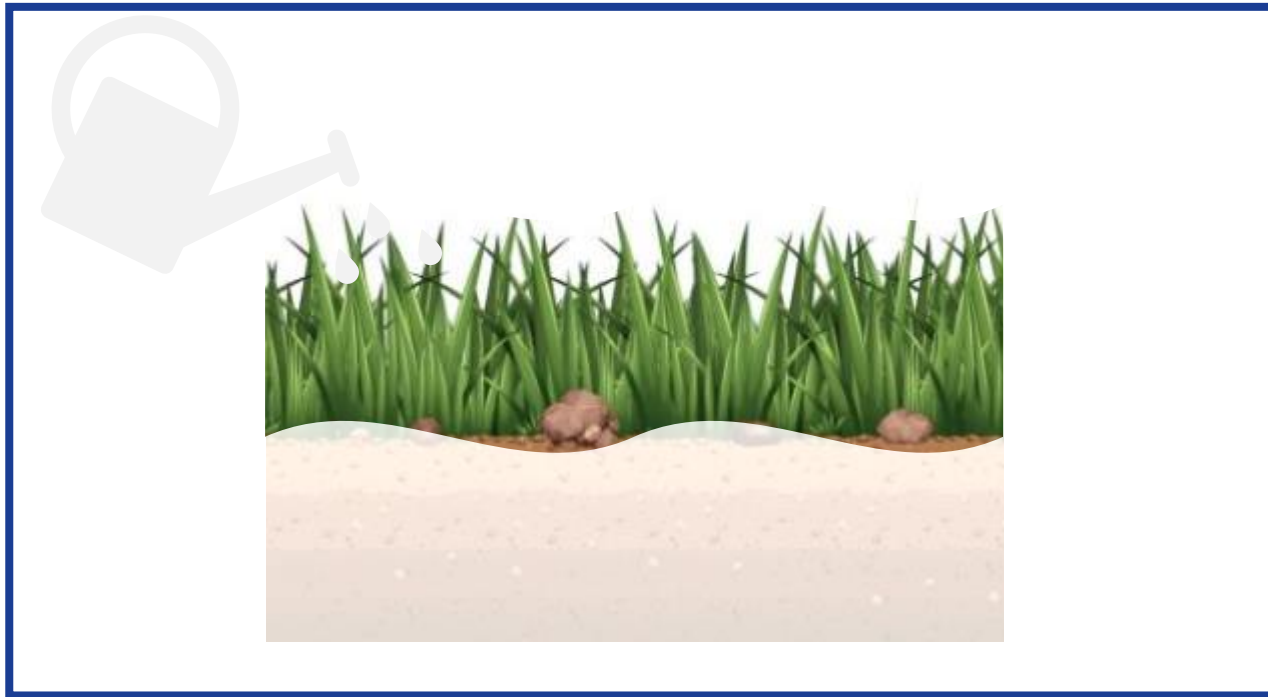
Conclusions

- Soil pollution reflects the area classification based on different levels of PFAS pressures: the highest concentrations were indeed found in samples from the red area.



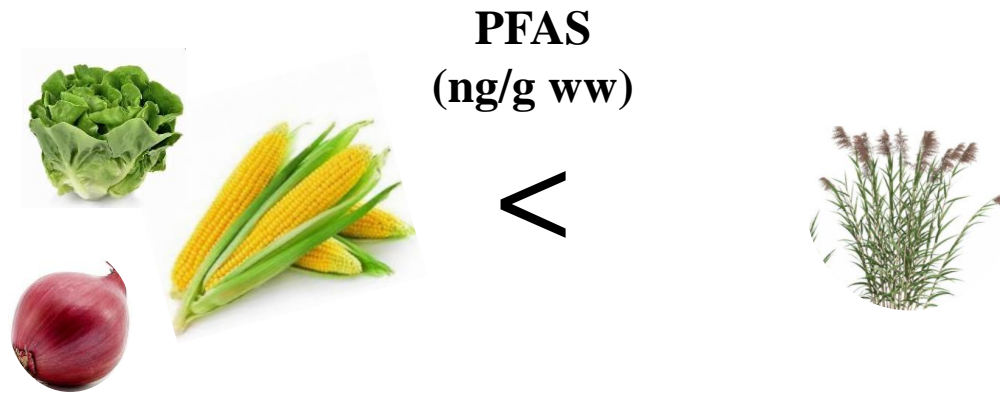
Conclusions

- In whole plants, contamination level was similar in the three areas, regardless of sampling site.
- Plants collected in the green area were mainly polluted by long-chain compounds ($C \geq 8$), released in the past, while short-chain chemicals ($C < 8$), of more recent origin, were dominant in vegetable samples taken in the red area, which is directly impacted by the industrial discharge in surface waters.



Conclusions

- PFAS concentrations in edible plant samples (onion, lettuce, corn kernels) were lower than those measured in spontaneous species and never exceeded 2 ng/g ww.



- The ability of reed grass to accumulate PFAS makes this plant the model organism in monitoring of contaminated areas
- Reed grass can be used also for phytodepuration in artificial wetlands

Thanks for your attention



Claudia Ferrario
IRSA-CNR Brugherio



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